



CARBOXY-THC IN HAIR BY LC-MS/MS OR GC-MS USING CLEAN SCREEN[®] THC EXTRACTION COLUMN

Part #

CSTHC206 – CLEAN SCREEN[®] THC 200 mg, 6 mL Tube

SMSTFA-1-1 – SELECTRA-SIL[®] MSTFA w/ 1% TMCS

SLDA50ID21-5UM – Selectra[®] DA HPLC Column, 50 x 2.1 mm, 5 μ m

1. PREPARE SAMPLE:

Into a clean glass tube add approx 100 mg of decontaminated hair
Add 1 mL of D.I. H₂O, add internal standard* and 100 μ L of 10 M NaOH
Digest at 70°C for 12 hours
Cool and adjust to pH 3

2. CONDITION CLEAN SCREEN[®] EXTRACTION COLUMN:

1 x 3 mL CH₃OH.
1 x 3 mL D.I. H₂O.
1 x 1 mL 100 mm HCl
NOTE: Aspirate at full vacuum or pressure

3. APPLY SAMPLE:

Load at 1 to 2 mL/minute.

4. WASH COLUMN:

1 x 3 mL D.I. H₂O
1 x 3 mL CH₃CN /HCl (30:70)
Dry column (10 minutes at full vacuum or pressure)

5. ELUTE THC-COOH:

1 x 3 mL Hexanes/ Ethyl Acetate/ Acetic Acid (49:49:2)
Collect eluate at 1 to 2 mL/minute

NOTE: Before proceeding, insure there are no water droplets at the bottom of the collection tube. This may increase drying time and decrease BSTFA derivatizing efficiency

6. DRY ELUATE:

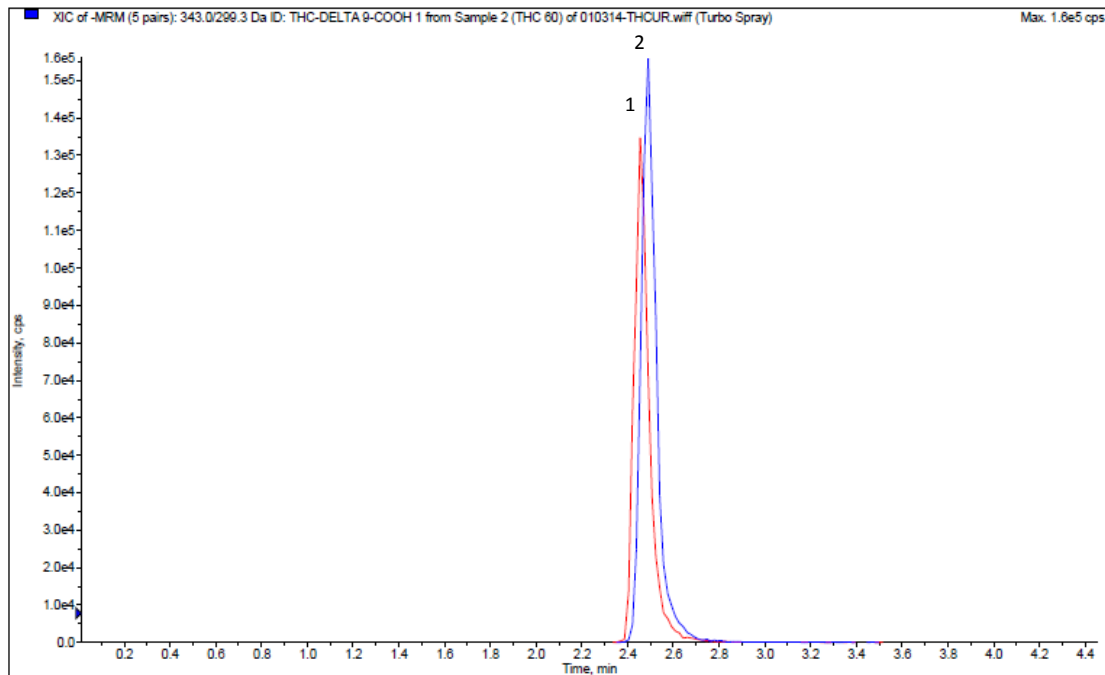
Evaporate to dryness at < 40 °C.

7. RECONSTITUTE:

Reconstitute sample in 100 μ L of mobile phase
Inject 20 μ L.

INSTRUMENT CONDITIONS (LC-MS/MS):

CHROMATOGRAM



Analyte	MRM Transitions		Relative Retention Time (min)
	Q1	Q3	
1. THC-DELTA 9-COOH D ₉	352	308	2.44
2. THC-DELTA 9-COOH	343	299	2.49

PARAMETERS

Mobile Phase A: 0.1% Formic Acid in D.I. H₂O

Mobile Phase B: 0.1% Formic Acid in Methanol

Flow Rate: 0.5 mL/minute

Polarity: Negative

Reconstitute: 100 µL

Injection Volume: 20 µL

LC Column: Selectra[®] DA HPLC Column 50 x 2.1 mm 5 µm

Instrument: API 3200 Qtrap MS/MS with Shimadzu Prominence UFLC

Gradient:

Time	%A	%B
0.00	60	40
2.00	30	70
2.50	10	90
2.51	60	40
4.00	STOP	