



## GHB IN URINE AND BLOOD WITHOUT CONVERSION TO GAMMA-BUTYRLACTONE (GBL) BY LC-MS/MS OR GC-MS CLEAN SCREEN® GHB EXTRACTION COLUMN

Part #

ZSGHB020 – CLEAN SCREEN® GHB 200 mg, 10 mL Tube

SBSTFA-1-1 – SELECTRA-SIL® BSTFA w/ 1% TMCS

### 1. PREPARE SAMPLE:

**Blood:** To 1 mL blood sample add internal standard and 0.5 mL of 100 mM phosphate buffer (pH 6.0).

Mix/vortex.

Rock for 10 minutes.

Centrifuge for 10 minutes at 2700 rpm.

**Urine:** To 200 µL of urine add internal standard(s) and 100 µL of 100 mM Phosphate buffer (pH 6.0)

Mix/vortex

### 2. CONDITION CLEAN SCREEN® EXTRACTION COLUMN:

1 x 3 mL CH<sub>3</sub>OH.

1 x 3 mL D.I. H<sub>2</sub>O.

1 x 1 mL 100 mM phosphate buffer (pH 6.0).

**NOTE:** Aspirate at full vacuum or pressure

### 3. APPLY SAMPLE:

Place test tubes into vacuum manifold for collection

The sample loading and elute are both collected

Load at 1 to 2 mL/minute

Collect elute as it is applied to the column

After the sample is off the columns apply full vacuum for about 15 seconds to remove any residual blood/urine.

### 4. ELUTE GHB:

Remove test tubes and set aside

Place clean tubes into vacuum manifold for collection

1 x 2 mL Methano/ NH<sub>4</sub>OH (99:1) onto SPE column and collect

### 5. DRY ELUATE:

Evaporate to dryness at < 40°C.

### 6. SAMPLE CLEAN-UP:

Add 200 µL of dimethylformamide.

Add 1 mL of hexane saturated with dimethylformamide.

Mix by inversion for 5 minutes.

Centrifuge at 3000 rpm for 5 minutes

Transfer lower dimethylformamide layer to a clean test tube

### 7. DRY ELUATE:

Evaporate to dryness at < 40 °C.

### 8. RECONSTITUTE / DERIVATIZE

- **LC-MS/MS:** Reconstitute sample in 100 µL of mobile phase  
Inject 20 µL

- **GC-MS: DERIVATIZE with TMS**

Add 50 µL Ethyl Acetate and 50 µL BSTFA w/ 1% TMCS

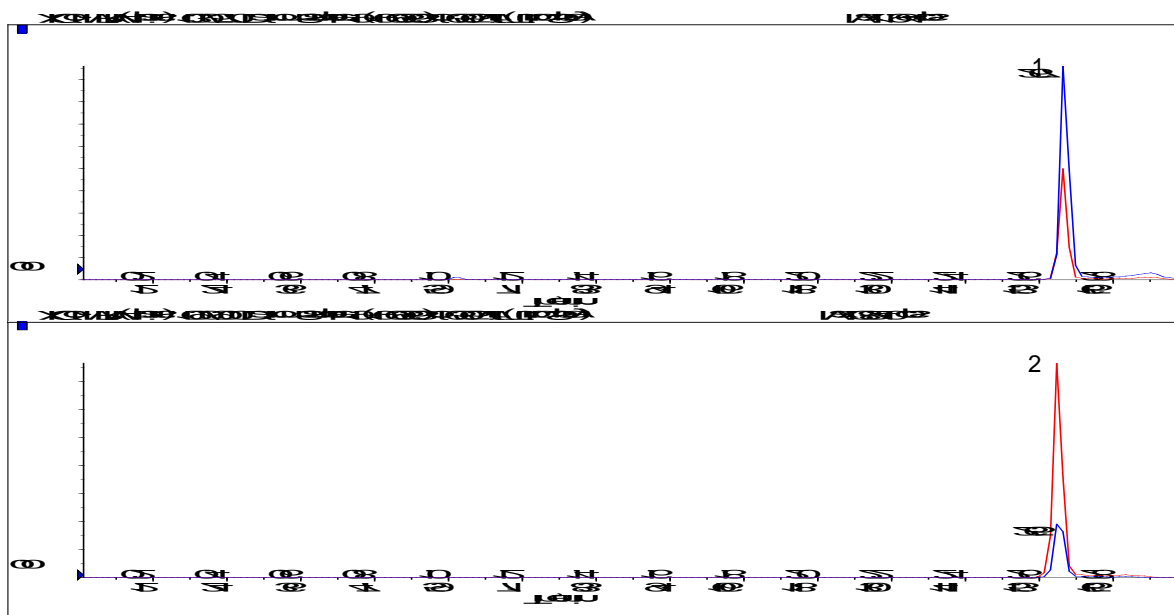
Overlay with N<sub>2</sub> and cap. Mix/vortex.

React 30 minutes at 70 °C. Remove from heat source to cool.

**NOTE:** Do not evaporate BSTFA solution

# INSTRUMENT CONDITIONS (LC-MS/MS):

## CHROMATOGRAM



Analyte	MRM Transitions		Relative Retention Time (min)
	Q1	Q3	
1.GHB	103.02	84.9	2.67
2.GHB-D <sub>6</sub>	109.13	90.0	2.65

## PARAMETERS

**Mobile Phase A:** 0.1% Formic Acid in D.I. H<sub>2</sub>O

**Mobile Phase B:** 0.1% Formic Acid in Acetonitrile

**Flow Rate:** 1.25 mL/minute

**Polarity:** Negative

**Reconstitute:** 100 µL

**Injection Volume:** 20 µL

**LC Column:** Biphenyl HPLC Column 150 x 4.6 mm 5 µm

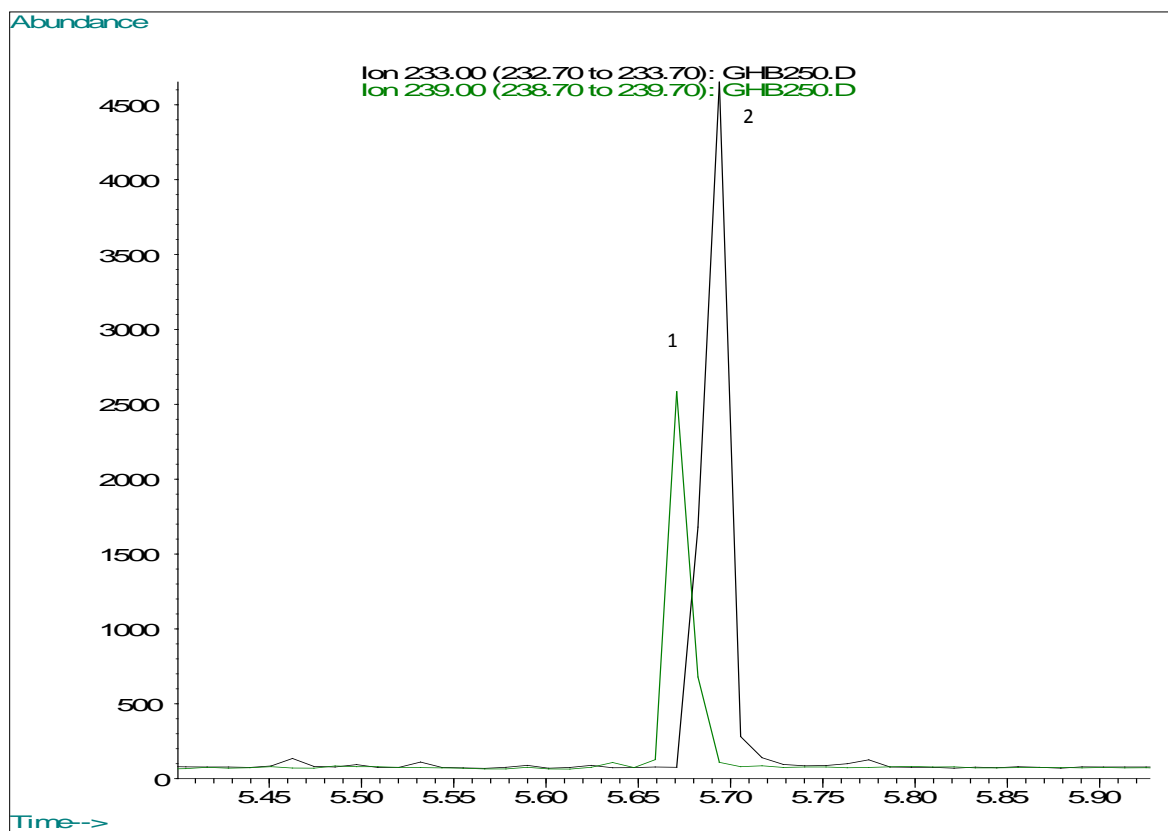
**Instrument:** API 3200 Qtrap MS/MS with Agilent 1200 Binary Pump SL

### Gradient:

Time	%A	%B
0.0	95	5
1.5	95	5
2.5	50	50
3.1	95	5
4.1	STOP	

## INSTRUMENT CONDITIONS (GC-MS):

### CHROMATOGRAM



### BSTFA-OXIME DERIVATIVES

Analyte	Quantify Ion	Qualifier Ion 1	Qualifier Ion 2	Relative Retention Time (min)
1. GHB-D <sub>6</sub>	239	240	241	5.67
2. GHB	233	234	235	5.69

### PARAMETERS

**GC/MS:** HP 5890 5972MSD GC/MS System with 7673 ALS System

**GC capillary column:** 30 m x0.25 mm (0.25 µm) RTX-5MS

**Injector:** 1µL Splitless 250 °C

**Oven temperature program:** 70 °C for 1 min; 15 °C/min to 130 °C, then to 300 °C 50 °C/min. Hold for 0.1 min

**Carrier gas:** Helium

**MSD condition:** Aux temperature: 280 °C, MS Source: 250 °C, MS Quad: 150 °C